



Editors: Basuki Supartono Ariyani Noviantari

Discovering the Miracle of Stem Cells

Discovering the Miracle of Stem Cells



Buku ini tidak diperjualbelikan.

First published in 2025 by BRIN Publishing Available to download free: penerbit.brin.go.id



This book is published under a Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International license (CC BY-NC-SA 4.0). This license allows you to share, copy, distribute and transmit the work for personal and non-commercial use providing appropriate attribution. Adaptation or modification to original work must be under the same license.

Further details about CC BY-NC-SA 4.0 licenses are available at https://creativecommons.org/licenses/by-nc-sa/4.0/

Editors: Basuki Supartono Ariyani Noviantari

Discovering the Miracle of Stem Cells



BRIN Publishing

Buku ini tidak diperjualbelikan.

© 2025 Editors & Authors

Cataloging in Publication

Discovering the Miracle of Stem Cells/Basuki Supartono & Ariyani Noviantari (Eds.)–Jakarta: Penerbit BRIN, 2025.

xxi p. + 3	393 p.; 14,8 x 21 cm.	
ISBN	978-602-6303-50-9	(e-book)
1. Stem ce	ells	2. Regenerative medicine
3. Stem ce	ell therapy	4. Tissue engineering therapy

616.15

Acquisition & Associate Ed	itor : Mayasuri Presilla
Copy editor	: Ayu Tya Farany & Muliyani
Proofreader	: Annisa' Eskahita Azizah & Martinus Helmiawan
Layouter	: Rina Kamila
Cover designer	: Rina Kamila
First Edition	: February 2025



Published by: BRIN Publishing, Member of Ikapi Directorate of Repositories, Multimedia, and Scientific Publishing Gedung B.J. Habibie Lt. 8, Jl. M.H. Thamrin No. 8, Kebon Sirih, Menteng, Jakarta Pusat, Daerah Khusus Ibukota Jakarta 10340 WhatsApp: +62 811-1064-6770 E-mail: penerbit@brin.go.id Website: penerbit@brin.go.id Penerbit BRIN @penerbit BRIN @penerbit_brin

Table of Contents

List of Figure	esix
List of Tables	sxiii
Publisher's N	lotexv
Foreword	xvii
Preface	xix
Chapter 1	Discovery of Stem Cells1 Ariyani Noviantari
Chapter 2	Stem Cell Culture Techniques: Mesenchymal Stem Cells
Chapter 3	Stem Cells for Orthopaedics Application49 Ismail Hadisoebroto Dilogo

Buku ini tidak diperjualbelikan.

Chapter 4	Mesenchymal Stem Cells Secretome for Diabetic Wound
Chapter 5	Mesenchymal Stem Cells and Their Conditioned Medium: For Skin Aging
Chapter 6	Stem Cells for Acute Myocardial Infarction: Safety and Efficacy
Chapter 7	Stem Cells Based Therapies for Neurological Disorders
Chapter 8	The Potential of Cd34+ Hematopoietic Stem Cell to Increase Fibroblast and Collagen Skin in Ultraviolet B-Exposed Skin
Chapter 9	Induced Pluripotent Stem Cells (iPSCs) and Neurological Diseases
Chapter 10	The Art of Ethical Dimensions in Stem Cell Research
Chapter 11	Stem Cells Are a New Hope, a New Horizon for Humanity and the Future of Human Beings: Representing Indonesia to the World

List of Abbreviations	299
Glossary	
About the Editors	351
About the Authors	355
Index	379

List of Figures

Figure 1.1	Potency of Differential of Stem Cells4
Figure 1.2	Differentiation Potential of MSCs
Figure 1.3	The Morphology of Primary MSCs Derived from Rat Bone Marrow
Figure 1.4	Anatomy of Umbilical Cord9
Figure 1.5	Sources of Stem Cells from Teeth11
Figure 2.1	Separation of MNCs Using a Separation Reagent27
Figure 2.2	Quantum Bioreactor
Figure 2.3	Plastic vessel attached explant culture facilitates "sprouting"
Figure 2.4	Simple Lipoaspirate Washing Using a Coffee Filter33
Figure 2.5	Flowchart of the Isolation of Adipose Tissue Derived Stromal Vascular Fraction
Figure 2.6	Schematic Picture of Umbilical Cord35
Figure 2.7	Schematic Picture of Multiple-Harvest Explants Method

Figure 4.1	Representative Image of Diabetic Chronic Ulcer Wound Treated with MSC CM83
Figure 6.1	LVEF examined by MRI and echocardiogram127
Figure 6.2	Quality of Life Assessment 129
Figure 7.1	Neurogenic niches of the brain include the subventricular zone (SVZ) and the subgranular zone (SGZ) of the dentate gyrus (DG)
Figure 7.2	Stages of neuronal development from neural stem or progenitor cells to mature neurons expressing biomarkers
Figure 8.1	Differences between Younger and Older Skin in Skin Structure
Figure 8.2	Mechanism of UV radiation mediates cellular damage
Figure 8.3	Markers of HSCs196
Figure 8.4	HSC Molecular Complexity Markers 197
Figure 8.5	HSC Molecular Development 200
Figure 8.6	HSC in Bone Marrow 203
Figure 8.7	HSC Hierarchy 204
Figure 8.8	HSC Lineage 207
Figure 8.9	Wistar Rat Dermal Tissue Fibroblast Cell Count with HE Staining
Figure 8.10	Collagen content in the dermis of male Wistar rats stained with Sirius Red
Figure 8.11	Differences in the Mean Number of Fibroblasts and Collagen between Groups after Administration of CD34+ Stem Cell Suspension
Figure 9.1	Direct Reprogramming of Somatic Cells into iPSCs 226
Figure 9.2	The Status of Human Pluripotence and Scit 227
Figure 9.3	The First Structure of the Human Brain Three Primary Cerebral Vesicles
Figure 11.1	Natural regeneration of cartilage defects in animal models results in fibrous tissue formation
Figure 11.2	Tissue Regeneration Cycle 275

Figure 11.3	Tissue Regeneration Cycle	276
Figure 11.4	Schematic Diagram of the Mechanism of Tissue Stem Cell Plasticity, from Waggers	281
Figure 11.5	Processing of the Tissue Stem Cells	284
Figure 11.6	Microscopic Images Of Tissue Regeneration (Oval Cells) in Naive Rats Following Repeated Intravenous Injections of Human PBMC Once a Month for Three Months	286
Figure 11.7	Safety of Administration of Peripheral Blood Mononuclear Cells (PBMC) to Patients with Unhealed Diabetic Wounds	287
Figure 11.8	Safety of Administration of Mesenchymal Stem Cells (MSCs) to Patients with Critical Bone Defects	288

List of Tables

Table 2.1	Final Concentration of Various Antibiotics and Antimycotics	44
Table 6.1	Summary of Case Illustration	125
Table 6.2	Regional Wall Motion Abnormalities (RWMA) and Wall Motion Score Index (WMSI) Result	131
Table 8.1	Histological Manifestations of Chronological Skin Aging	188
Table 8.2	Clinical Trials Applications of Stem Cell for Facial Skin Aging and Photoaging	205
Table 9.1	Summary of Neurological Disease Modelling with iPSC	236

Preface

The journey into stem cell research represents one of the most profound scientific endeavors of our time. From the early exploration of basic cell biology to the groundbreaking advances in regenerative medicine, stem cells have opened up new possibilities for healing and restoring the human body. *Discovering the Miracle of Stem Cells* seeks to highlight these extraordinary potentials, particularly focusing on the contributions made by Indonesian researchers to this rapidly evolving field.

This book covers a wide array of topics within stem cell science, encompassing both fundamental research and clinical applications. It underscores the pivotal role that stem cells have played in transforming modern medicine. The initial chapters, such as "Stem Cell Culture Techniques" by Jeanne Adiwinata Pawitan, provide key insights into culturing methods and the development of mesenchymal stem cells (MSCs), which are among the most widely applied stem cells in regenerative therapies. Clinical applications are thoroughly explored in chapters like "Stem Cell for Orthopaedics Application" by Ismail Hadisoebroto Dilogo, as well as discussions on the potential use of stem cells in treating diabetes and skin aging, with contributions from experts such as Siufui Hendrawan and Winawati Eka Putri. These chapters are enriched with fascinating case studies in orthopaedics, cardiology, dermatology, and neurology, offering a comprehensive overview of how stem cells are being utilized in diverse medical fields.

One notable chapter, "The Potential of CD34+ Stem Cells in Increasing Fibroblast and Collagen Levels in Ultraviolet B Exposed Skin," co-authored by Basuki Supartono, explores the use of stem cells in anti-aging therapies, providing promising avenues for skin regeneration. In addition, the book delves into the ethical and legal dimensions of stem cell research, expertly analyzed by Dito Anurogo in the chapter "The Art of Socio-ethical and Legal Dimensions in Stem Cell Research." The articles in this book have been carefully selected, reviewed, edited, and proofread under the supervision of the National Research and Innovation Agency (BRIN), ensuring the highest level of academic integrity.

On a broader scale, this book not only consolidates existing knowledge in the field of stem cells, but also introduces new perspectives and clinical applications specific to the Indonesian healthcare context. While global research on stem cells is vast, this work offers a unique focus on Indonesia's advancements, illustrating how local research and clinical trials are contributing to global progress. By situating these developments alongside international breakthroughs, *Discovering the Miracle of Stem Cells* adds a regional dimension to the existing body of literature in regenerative medicine, offering insights into localized challenges and solutions often overlooked in mainstream research.

As a researcher and clinician, I have had the privilege of witnessing firsthand the remarkable potential of stem cell therapies to provide hope and relief to patients suffering from a wide range of conditions.

I am deeply thankful to my colleagues, research teams, and collaborators, whose unwavering dedication and tireless efforts have made this book possible. Their contributions have been instrumental in advancing our collective understanding of stem cells and propelling this pioneering field to the forefront of medical innovation. I also extend my sincere gratitude to the National Research and Innovation Agency (BRIN) for their invaluable support.

I offer my heartfelt thanks to the readers of this book, particularly practitioners, researchers, and students in the biomedical sciences. My hope is that this book will serve both as a valuable reference and a source of inspiration, encouraging further research and innovation in stem cell science.

Lastly, I wish to acknowledge the steadfast support of my wife, Dr. Prita Kusumaningsih, SpOG, my family, my children, grandchildren, and friends, who have continuously encouraged me throughout the writing process. Their patience and understanding have been invaluable in bringing this work to completion.

As we continue to explore the vast potential of stem cells, I believe we are only beginning to realize the full scope of their capabilities. *Discovering the Miracle of Stem Cells* is not merely a record of scientific discoveries; it is a testament to the future of medicine, where healing and regeneration become achievable for all.

Jakarta, October 12th, 2024

Prof. Dr. dr. Basuki Supartono, Sp.O.T., F.I.C.S., M.A.R.S

Chapter 4

Mesenchymal Stem Cells Secretome for Diabetic Wound

Siufui Hendrawan Jennifer Lheman David Victorious Lukas Sukmawati Tansil Tan

A. Diabetes and Its Complications

Diabetic complications can be generalized into two categories, microvascular and macrovascular complications. Microvascular complications are related to complications affecting microvessels such as retinopathy, neuropathy, and nephropathy. On the other hand, macrovascular complications are related to those affecting macrovessels such as coronary artery disease, peripheral artery disease, and stroke (Mauricio et al., 2020; Ohiagu et al., 2021). The macrovascular complications, such as coronary artery disease, had a high prevalence among people suffering from diabetes. Besides that, one of the most common complications of diabetes is peripheral artery disease, which is associated with other diseases, particularly diabetic

Tarumanagara University, e-mail: siufui@fk.untar.ac.id

Hendrawan, S., Lheman, J., Lukas, D. V., & Tan, S. T. (2025). Mesenchymal stem cells secretome for diabetic wound. In B. Supartono & A. Noviantari (Eds.), *Discovering the miracle of stem cells* (73–95). BRIN Publishing. DOI: 10.55981/brin.1128.c1298, E-ISBN: 978-602-6303-50-9

S. Hendrawan, J. Lheman, D. V. Lukas, S. T. Tan

^{© 2025} Editors & Authors

peripheral neuropathy. Both these disorders could lead to diabetic foot which usually leads to nonhealing foot ulcers (Mauricio et al., 2020; Ohiagu et al., 2021). There are still many diabetic complications other than those mentioned above.

B. Underlying Mechanism of Diabetic Complications

According to Shi et al. (2018), several studies have shown that hyperglycemic conditions may affect microvascular and macrovascular disruptions. This disease is a major contributor to the development of vascular problems and can mediate the negative effects through many mechanisms (Shi et al., 2018). The mechanisms include increased oxidative stress, mitochondrial dysfunction, excessive cellular apoptosis, and abnormal cellular autophagy, which is caused by long-term diabetic condition and have a potential effect on diabetes complications, whether it is directly or indirectly (Shi et al., 2018). In brief, this excessive production of ROS will increase the synthesis of strong oxidative peroxynitrite, induced damage to DNA, and increased the risks of diabetic microvascular complications throughout several mechanisms (Shi et al., 2017, 2018). The next mechanism is related to cellular and tissue injury in diabetes is cellular apoptosis. Apoptosis is a mechanism for genetically programmed cell death, and it has a major role in the survival of an organisms (Hamzawy et al., 2017). Cell apoptosis related to diabetes is mediated by two substantial forms: stress on endoplasmic reticulum and damage to mitochondria. There are two key pathways that take part in this mechanism, the extrinsic death receptor pathway and intrinsic mitochondrial pathway (Huang et al., 2017; Shi et al., 2018). Another major mechanism involved is autophagy which is another significant way to maintain environmental homeostasis in intracellular (Hamzawy et al., 2017). Being in a hyperglycemic condition for a prolonged period of time, abnormal cellular autophagy could occur. This mechanism is activated when the cell condition is in various stress situations such as lack of essential nutrients and high glucose environment. Autophagy is necessary for cellular functions, the deficiency of this mechanism could lead to cellular degeneration and disruption of intracellular homeostasis (Hamzawy et al., 2017; Shi et al., 2018). Apart from the mechanisms mentioned above, there are some that believe that growth factors play a role in the development of abnormal growth and impaired regeneration in diabetics (Shi et al., 2018). Thus, it is important to understand the underlying mechanism that is impaired due to this condition.

C. Diabetic Ulcer as One of Serious Concern Complications of Diabetes

Diabetic ulcers are arising as a serious concern due to the complications from amputation cases. The incidence rate of minor and major amputations related to diabetes was 139.97 and 94.82 events respectively, per 100,000 diabetic patients per year. In the case of lower extremity amputations related to diabetes, it is more than two-fold higher in patients with type 1 DM than those with type 2 DM (Ezzatvar & García-Hermoso, 2022). This condition is a result of disruption in all phases of wound healing as a result of diabetes itself (Okonkwo et al., 2020). There are numerous factors that are responsible for the impaired wound healing process in diabetic patients, including impaired growth factor production, angiogenic response, and collagen formation (Fui et al., 2019). Consequently, even slight skin abrasions or scratches tended to develop into chronic wounds in diabetic patients (Avishai et al., 2017).

1. Impaired Wound Healing Process in Diabetes

Normally the wound healing process can be divided into four phases, specifically hemostasis, inflammation, proliferation, and finally the remodeling process (Fui et al., 2019). In brief, hemostasis is the first phase of the process in which the constriction of blood vessels occurs and platelet cells aggregate to clot the wound. The next phase is the inflammatory stage, when inflammatory cells migrate to the wound and blood circulation is increased (4 to 6 days). Then, proliferative

stage, which is when the wound bed is filled with granulation tissue during this phase, followed by angiogenesis which is when the new blood vessels are formed (4 to 21 days). Epidermal cells such as fibroblasts and keratinocytes are proliferated and migrated across the wound. After that, the collagen accumulation happens, then finally the wound edges are contracted. The final phase is remodeling phase which occurs 21 days to 2 years post injury. The tensile strength is increased by collagen crosslinking and the scar is matured (Fui et al., 2019). If there are one or more of these phases impaired, consequently the wound healing process becomes delayed and it will lead to chronic ulcer development.

In diabetic patients with a constant state of hyperglycemia, the endothelial cells lose their integrity and eventually become susceptible to apoptosis and detachment, resulting in impairment of the wound healing process (Okonkwo & Dipietro, 2017; Velnar & Gradisnik, 2018). The study conducted by Okonkwo et al. (2020) also found that there was a decreased amount of functional endothelial cells present in the diabetic skin. There was also impairment in the pruning and refinement process of the capillary bed (Bodnar et al., 2016; Caporali et al., 2017; Okonkwo et al., 2020). Therefore, it leads to the conclusion that skin wounds on diabetic patients are significantly reduced due to neovascularization and pro-angiogenic factor expression after injury (Okonkwo et al., 2020). Moreover, the overproduction of ROS in diabetics will result in cellular damage of endothelial cells (Fui et al., 2019).

Although many comprehensive treatments are currently available, the number of complications related to diabetic wounds, especially diabetic foot ulcers, is still alarming, with the data reported by the IDF showing that 9.1–26.1 million people will develop diabetic foot ulcers (Armstrong et al., 2017; Everett & Mathioudakis, 2018). Hence, advanced treatments for diabetic wounds have been studied intensively in the last decade. Growth factors are known to be involved in every phase of wound healing through their inhibitory or stimulatory effect (Burgess et al., 2021; Fui et al., 2019). Although there is some belief that in the diabetic condition, growth factors play a major role in repairing tissue, nevertheless, it is still one of the promising targets for diabetic wound healing therapy.

2. Growth Factors That Related to Wound Healing Process

There are some major growth factors that play an important role in the wound healing process. Platelet-derived growth factor (PDGF) is released by platelets as a key factor that functions to increase the infiltration of immune cells, activating macrophages, promoting fibroblast proliferation, and accumulation of the extracellular matrix (ECM). It is involved in the inflammatory, proliferative, and remodeling phases of wound healing (Gardner et al., 2016; Patel et al., 2019). Epidermal growth factor (EGF), which is involved in the proliferation phase, is also released by platelets. It has the function of enhancing cell motility, migration, and cell proliferation (Bai et al., 2016; Fui et al., 2019; Patel et al., 2019; Shin et al., 2022). Transforming growth factor beta 1 (TGF- β 1), involved in inflammatory and proliferation phase, it is produced by several cells such as macrophages, fibroblasts, keratinocytes, and platelets. The functions of TGF-\u00df1 are increasing leukocytes and fibroblast migration, promoting angiogenesis, and also stimulating the production of ECM components (Bai et al., 2016; Fui et al., 2019; Mori et al., 2016). Vascular endothelial growth factor (VEGF) is the most known growth factor for its potent angiogenic properties. It increases capillary density and improves the blood metabolism in wounded tissue. It also mediates migration and proliferation of endothelial cells, angiogenesis and tissue granulation during inflammatory and proliferation phases (Fui et al., 2019; Gardner et al., 2016; Patel et al., 2019). Finally, basic fibroblast growth factor (bFGF) is usually highly expressed during late inflammatory stage. This growth factor has the functions to enhance the proliferation of fibroblast, promotes angiogenesis and collagen maturation during the proliferation and remodeling phase of the wound healing (Bai et al., 2016; Fui et al., 2019). Most of these growth factors are secreted by

mesenchymal stem cells, which is why it has become the major focus for advanced therapy in diabetes.

D. Stem Cell-Based Therapy for Diabetes

Recently, stem cells-based therapy has arisen as one of promising alternative treatment to many diseases, including diabetes. Interestingly, mesenchymal stem cells (MSCs) are well known for their regenerative ability and immunomodulatory attributes. MSCs can be found in many perivascular tissues such as bone marrow, adipose tissue, teeth, placenta, umbilical cord, amniotic fluid, and cord blood (Shin et al., 2021). The following are some studies, both pre-clinical and clinical trials, that used mesenchymal stem cells from varied sources to treat diabetic condition such as insulin resistance and control the hyperglycemia condition.

Wharton's jelly derived MSCs and hematopoietic stem cells derived from umbilical cord blood have been analyzed for treatment of type 1 and type 2 diabetes (Bani Hamad et al., 2021). A clinical trial in China used Wharton's jelly derived MSCs on a recently diagnosed type 1 DM patients. This clinical trial was designed as randomized controlled study, and the stem cells were administered via intravenous injection and were combined with insulin administration prior and throughout the follow up period. The dose of implanted MSCs was not disclosed by the authors. The result showed improvement in hemoglobin A1c (HbA1c) levels on the stem cells therapy group, the dosage of insulin administration was significantly decreased, and interestingly, a fifth of patients in the therapy group become insulinindependent, for almost 1.5 years. Moreover, there were no adverse events reported during the study (Bani Hamad et al., 2021; Hu et al., 2013). Other clinical trial was done on type 2 DM patients in China. Wharton's jelly derived MSCs is used and administered twice. First, it was delivered via intravenous injection, then for the second dose it was directly injected through splenic artery using catheter. The first and second dosage was given five days apart. The result showed

there was a decrease in HbA1c and blood glucose levels, moreover the dosage of insulin and other anti-diabetic medication is reduced. Although there are no control group to compare the result in this trial, this result showed that administration of Wharton's jelly derived MSCs can improve the regulation of metabolic pathway and β cell function in type 2 DM patients (Bani Hamad et al., 2021; Liu et al., 2014). The limitation for both studies is the small sample size. Overall, it proved that stem cell therapy was much safer compared to islet and organ transplantation (Bani Hamad et al., 2021). Umbilical cord (UC) derived MSCs are considered a better choice for clinical applications due to its high paracrine potential and it has low immunogenicity (Wang et al., 2018; Xiang et al., 2020). MSC have the ability to repair the cell damage through paracrine mechanisms from several factors such as immunomodulation factors, angiogenic factors, antiapoptotic factors, antioxidative factors, and also cell migration, and targeting and stimulation, although their fundamental and detailed biological mechanism still required further elucidation (Gnecchi et al., 2016; Liang et al., 2014).

Despite its promising results, there are some limitations of this stem cells therapy. These are related to the administration of stem cells, which is via infusion route in most of the studies or via clinical trials regarding type 1 or type 2 DM (Cho et al., 2018). One of the significant challenges is the low survival rate of the engrafted cells. Many transplanted cells eventually will die within hours or days post transplantation (Mitrousis et al., 2018; Sortwell et al., 2000). Numerous efforts have been attempted by researchers to overcome this problem. Pre-conditioning, genetic modification, and mimicking extracellular matrix such as hydrogel have been used to improve the survival of the cells (Li et al., 2016; Zhao et al., 2019). Nonetheless, further strategies and research are needed, including various cells condition and environment, the delivery system, and dosages that must be consider.

E. Secretome Based Therapy for Diabetes

This last decade, researchers were racing to develop cell free therapy derived from stem cells. As described above, stem cells, such as MSCs, have been used in clinical trials to treat diabetes and there are several trials that have proven their positive effects. However, due to some challenges to have the optimal effect from the usage of MSCs, cell-free therapy such as secretome or CM is more preferrable. MSCs from various sources have been known to release numerous paracrine factors that classified as bioactive molecules (Hsiao et al., 2011). These bioactive molecules which secreted into the extracellular space are known as secretome and is secreted by MSCs as a response to specific microenvironment conditions. According to González-González et al. (2020), secretome contains two different components. The first component is a soluble part, mostly comprised of cytokines, chemokines, immunomodulatory molecules, and broad spectrum of growth factors (Madrigal et al., 2014). The second component is a vesicular fragment, consisted of variety type of extracellular vesicles (EVs) (González-González et al., 2020; Teixeira & Salgado, 2020) such as microvesicles (Bruno et al., 2009), microparticles (Kim et al., 2012) and exosomes (Lai et al., 2010, 2015). Various studies on these secreted factors showed that even without the cells, it still has the regenerative ability to repair tissue or organ damage (Pawitan, 2014). This secreted factor can be found in the media where the cells were cultured, therefore the media is called conditioned medium (CM) (Kim et al., 2013).

There are some theories stating that the origin of the MSCs may have a difference in the protein expression. A study conducted by Shin et al. (2021) has done the comparative analysis of the secretome from different sources which are adipose and bone marrow (adult stem cells) and placenta and Wharton's jelly (fetal stem cells). There were plenty of proteins that involved in cellular migration and apoptosis reduction in the secretome derived from adipose, placenta, and Wharton's jelly, but not from bone marrow, though the level is varied between the sources. On adult stem cells, protein secreted by adipose MSCs is associated with the organization such as the development of cytoplasm, while protein secreted by bone marrow MSCs is related to cellular development, and epithelial-mesenchymal transition. Protein that associated to cell migration and survival were detected similarly on both sources (Shin et al., 2021). Nonetheless, secretome secreted by fetal MSC group was expected to have higher potential than the adult stem cells due to the higher quantity of protein and more diverse proteins they have (Shin et al., 2021).

Secretome or CM has been used in several pre-clinical research particularly for wound healing in diabetes case. In brief, these are some of the studies related to that case. First is an in vitro study using secretome isolated from human adipose tissue-derived MSC has showed its ability to accelerate cutaneous wound healing. The results showed that the epidermal and dermal thickness, vascularized granulation tissue, and dermal collagen layers were increased on the wound treated by the stem cell secretome. The secretome stimulates collagen synthesis and migration of dermal fibroblasts through upregulating the transcription of collagen type I and III. It also may promote wound healing by increasing re-epithelization of the dermal tissue (Park et al., 2018). MSC and MSC-CM accelerated epithelialization, increasing granulation tissue formation. In response to MSC and MSC-CM, dermal fibroblast secrete increased the amounts of collagen type I and alter gene expression (Gnecchi et al., 2016).

We also have done an in vivo study using conditioned medium isolated from human umbilical cord-derived MSCs in diabetes induced rats to observe the wound healing potential. All animal experiments in this study were approved by Institutional Animal Care and Use Committees (IACUC) of the Faculty of Medicine, Tarumanagara University, approval number 001.KEPH/UPPM/FK/IV/2019. The cells were processed at Stem Cell and Cancer Institute Laboratory, Jakarta, Indonesia. The MSCs were cultured under hypoxic condition then the CM was collected. The result showed that this pre-conditioning hypoxic condition could stimulate MSCs to produce higher growth factors such as VEGF, bFGF, and pro-collagen 1 and promote better wound closure in rats. Intriguingly, VEGF was not secreted in CM collected from umbilical cord MSCs that was cultured in normoxic condition. It has been proven that pre-conditioning certain factors such as hypoxia could enhance growth factors secretion. The histopathological analysis on the wound site showed that there is an increase in re-epithelization and also has the largest collagen deposition in the group treated using hypoxic umbilical cord-CM compared to the other group. Therefore, we concluded that the CM collected from umbilical cord MSCs cultured in hypoxic condition has positive effects towards wound healing process based on the result of re-epithelization and collagen formation on the wound site (Hendrawan et al., 2021).

Another in vivo study was done by Saheli et al. (2020) which evaluated the impact of CM collected from human bone marrow derived MSCs for diabetic wound healing in rats. The result showed the healing progress on the diabetic wound treated by CM was improved and comparable to the progress on non-diabetic group. They also found that the inflammation was significantly reduced on day 4 in the group treated with the CM compared to the diabetic control group. Higher expression of EGF and bFGF was also observed on the diabetic wound treated by the CM. Additionally, the collagen density was also increased, the inflammation was repressed, number of fibroblasts and microvessels was significantly elevated on the CMtreated group when compared to the diabetic wound. Therefore, this study also demonstrated that administration of MSC-CM has the potential to effectively improve the quality of healed wounds in chronic diabetes condition, which mainly through the modulation of fibroblast behaviors (Saheli et al., 2020).

Even though numerous researches were done using secretome or CM in relation to wound healing, especially diabetic wound, however, there are still only a few pre-clinical trials using CM or secretome systemically to treat hyperglycemic condition and other diabetic complications. We have done a pilot study to see the effect using CM from hypoxic human umbilical cord MSCs via intravenous injection on diabetic induced rats. The pilot study was approved by IACUC of PT. Bimana Indomedical, Bogor, ethical approval number R.02-21-IR. The CM was injected intravenously through rat tail vein and the blood glucose concentration was monitored for 1 month. The results showed that the insulin concentration was decreased in CM group and was comparable to the normal rats. Based on this result, it suggested that the administration of CM could reduce the hypersensitivity of β cells. There were no side effects observed during the study (unpublished data) (Hendrawan et al., 2021; Tan et al., 2021).

We also performed a clinical study (number NCT04134676), in which we evaluated the therapeutic potential of CM on chronic ulcer wounds especially on diabetic patients. The ethical clearance for this study was obtained from Human Research Ethics Committee, Institute of Research and Community Engagement of Tarumanagara University, number 1007-Int-KLPPM/Untar/VI/2020. Umbilical



Notes: (A) Before treatment and (B) After treatment Photo: Sukmawati Tansil Tan (2021) Figure 4.1 Representative Image of Diabetic Chronic Ulcer Wound Treated with MSC CM

83

cord was obtained with the parental consent and was processed at Tarumanagara Human Cell Technology Laboratory, Jakarta, Indonesia. The CM was collected from MSC cultured under hypoxic condition (Figure 4.1a). The CM and other active ingredients were mixed in the form of 10% gel for topical use. An ample amount of the gel was applied to the wound. The results showed that the width and length of the wound decreased. Moreover, the bed of wounds is improved after 2 weeks post treatment (Figure 4.1B). The wound was treated for one month. There are no adverse effects observed in this study. Overall, the study showed that the topical administration of 10% gel CM can effectively enhance wound healing, in particular diabetic chronic ulcers (Tan et al., 2023).

F. Future Prospect and Challenge Against Secretome Wide Clinical Application

Stem cell-based therapy has emerged as a prominent alternative therapy to various degenerative diseases (Park et al., 2018). However, there are numerous challenges for clinical application of this therapy. The process of manufacture, sources, cell culture protocols, level of expansion and status of the cells can influence the therapeutic effectiveness of MSCs. Until now, allogenic or autologous MSCs have been used in clinical trials, while xenografts of MSCs are only applied in pre-clinical studies (Shin et al., 2021). The concerns are delivery route and dosage of the MSCs. Viable cells usually were delivered to the body via injection or catheter. However, the data showed that injection of live cells through a syringe needle can reduce the cell viability below 32% and it could cause irreparable damage to the cell membrane. It could also lead to a raise of an immune response that can be harmful for healing process (Ahangar et al., 2020). Moreover, there are side effects that are associated to MSC administration such as transient fever, constipation, and fatigue. Neither serious adverse events nor mortality were discovered across the clinical studies (Wang et al., 2021).

On the contrary, secretome provides an option for cell-free therapy with lower immunogenicity reaction. Secretome has the advantage that it can be prepared ahead in larger quantities and immediately become available for application (Xia et al., 2019). Despite the fact that it is easier to produce, handle, and store compared to the viable cells, it also has several drawbacks and challenges to bring it to bedside application (González-González et al., 2020).

To date, there is still no clinical trials have been registered in the clinicaltrials.gov that used the mesenchymal stem cells secretome or conditioned medium, whether to treat diabetic condition such as to control the hyperglycemia or to treat the diabetic complications cases. In brief, here are some of the challenges with secretome especially secretome from MSCs as therapeutic product. First, the characterization of secretome is needed. Due to the composition of secretome, it has become highly challenging to define specific function of each components and quantify the activity (Ahangar et al., 2020; Vizoso et al., 2017). Secondly, the inconsistency during the preparation of secretome from the MSCs. It is well-known that there are many factors that could affect the quality and efficacy of the secretome. Health condition and age of the donors, also the methods for isolation and culture the MSCs, are some of the factors that must be considered. The donor should be strictly screened and free from hepatitis B virus and human immunodeficiency virus (HIV) (Ahangar et al., 2020; Lukomska et al., 2019). The source of the MSCs is also one of the hurdles, some of it possibly due to ethical issue. For example, the usage of human fetal which obtained from the abortion procedure, although it has unique properties, has ethical issue for clinical application. Conversely, human umbilical cord should be more suitable as source of MSCs because it was categorized as clinical waste, therefore, there are no ethical issue to use the umbilical cord tissue (Wang et al., 2023). The other challenges regarding inconsistency are the heterogenicity of the MSCs, number of cells, and the interval of time. The most crucial part of the challenges is to produce the secretome under pharmaceutical standard and in the Good Manufacturing Practice (GMP) certified

facility. The secretome production under good manufacturing protocols can improve the consistency from one batch to another and importantly the efficacy of the secretome can be reproducible (Ahangar et al., 2020; De Sousa et al., 2016). The other concern part of secretome application is the potential side effect of the secretome administration. Despite the fact that there are only a few reports regarding the negative effects or even adverse events of secretome, there is always a risk that potentially happens when administering foreign substance. One of the problems is the immunosuppressive properties that has been reported in some studies (Zhao et al., 2016). Therefore, there is probability that the usage of secretome could cause immunodeficiency and poses risk to an infection (Bascones-Martinez et al., 2014). Hence, the optimum dosage for secretome administration should be clearly specified to have the balance of efficacy and safety of this secretome based treatment (Ahangar et al., 2020). Another concern regarding the instability and half-life of the protein contained in the secretome could be overcome by pre-conditioning the cells to increase the paracrine activity and production of the cells (Park et al., 2018). The alteration, namely hypoxia, inflammatory stimulus, or even the usage of bioreactors on preconditioned cells, was also reported to be related to increases the therapeutic potential of secretome (Pinho et al., 2020). We have proved that pre-conditioning such as culturing the cells in a hypoxic condition will increase the growth factor production in the secretome compared to the cells culture in normal condition (Hendrawan et al., 2021). Besides the challenges, there are concerns such as the route of administration, dosage, and duration of secretome application that need to be determined and standardized. The most common route of administration is topical for wound healing treatment (Fui et al., 2019). For other therapy, there is pre-clinical trial that used injection (intramuscular, intravenous) as the delivery route. However, there is still no clinical trials for diabetes that administered secretome as its therapy. Related to dosage, it will become one of the difficult challenges to calculate the generalized

dosage and duration needed for diabetes related disease. Although there is a study that has demonstrated the repeated administration of secretome can increase the duration of secretome effects as we have described above, the available data is still very limited. Furthermore, for wider clinical application, it is necessary to apply the precautionary principle and based on scientific evidence on safety and efficacy. It is also necessary to increase the education of the general public on how to interpret and apply these new findings. In Indonesia, the support from the government is obvious and have already regulated the provision of stem cell services which has all been stated in Indonesian Minister of Health Regulation Number 32, 2018 (Permenkes No. 32, 2018). However, there are still limited GMP certified facilities that has been established in the country. Limited budget is also one of the biggest concerns, which is why there is still a long way to go to bring the secretome to bedside application widely in Indonesia. Hopefully, the government concern to decrease the number of diabetes related complications could ease the way of secretome clinical application in Indonesia. In conclusion, there are numerous treatments for diabetes; nonetheless, the complications of this disease are still happening.

There are evidences that in most of the pre-clinical and clinical trials that used stem cell-based therapy which showed positive results, especially regarding diabetic wound healing treatment. While there are many challenges, this therapy is highly potential and very promising as alternative therapy for diabetes and its complications in translational medicine. New strategies are needed for overcoming limitations of stem cells and its secretome to be applied in a broad field of disorders, such as the use of encapsulated stem cells (Freimark et al., 2010) and nanotechnology (Zaghary et al., 2021). Moreover, cooperation among all stakeholders is essential to accelerate clinical applications. Good clinical trials to prove safety and actual efficacy of stem cell therapy are required to rush application and development of commercialized products.

87

References

- Ahangar, P., Mills, S. J., & Cowin, A. J. (2020). Mesenchymal stem cell secretome as an emerging cell-free alternative for improving wound repair. *International Journal of Molecular Sciences*, 21(19), Article 7038. https://doi.org/10.3390/ijms21197038
- Armstrong, D. G., Boulton, A. J. M., & Bus, S. A. (2017). Diabetic foot ulcers and their recurrence. *New England Journal of Medicine*, 376(24), 2367–2375. https://doi.org/10.1056/nejmra1615439
- Avishai, E., Yeghiazaryan, K., & Golubnitschaja, O. (2017). Impaired wound healing: Facts and hypotheses for multi-professional considerations in predictive, preventive and personalised medicine. *EPMA Journal*, 8, 23–33. https://doi.org/10.1007/s13167-017-0081-y
- Bai, L., Li, D., Li, J., Luo, Z., Yu, S., Cao, S., Shen, L., Zuo, Z., & Ma, X. (2016). Bioactive molecules derived from umbilical cord mesenchymal stem cells. *Acta Histochemica*, 118(8), 761–769. https://doi.org/10.1016/j. acthis.2016.09.006
- Bani Hamad, F. R., Rahat, N., Shankar, K., & Tsouklidis, N. (2021). Efficacy of stem cell application in diabetes mellitus: Promising future therapy for diabetes and its complications. *Cureus*, 13(2), Article e13563. https://doi.org/10.7759/cureus.13563
- Bascones-Martinez, A., Mattila, R., Gomez-Font, R., & Meurman, J. H. (2014). Immunomodulatory drugs: Oral and systemic adverse effects. *Medicina Oral, Patologia Oral y Cirugia Bucal*, 19(1), 24–31. https:// doi.org/10.4317/medoral.19087
- Bodnar, R. J., Satish, L., Yates, C. C., & Wells, A. (2016). Pericytes: A newly recognized player in wound healing. *Wound Repair and Regeneration*, 24(2), 204–214. https://doi.org/10.1111/wrr.12415
- Bruno, S., Grange, C., Deregibus, M. C., Calogero, R. A., Saviozzi, S., Collino, F., Morando, L., Busca, A., Falda, M., Bussolati, B., Tetta, C., & Camussi, G. (2009). Mesenchymal stem cell-derived microvesicles protect against acute tubular injury. *Journal of the American Society of Nephrology*, 20(5), 1053–1067. https://doi.org/10.1681/ASN.2008070798
- Burgess, J. L., Wyant, W. A., Abujamra, B. A., Kirsner, R. S., & Jozic, I. (2021). Diabetic wound-healing science. *Medicina*, 57(10), Article 1072. https://doi.org/10.3390/medicina57101072
- Caporali, A., Martello, A., Miscianinov, V., Maselli, D., Vono, R., & Spinetti, G. (2017). Contribution of pericyte paracrine regulation of

the endothelium to angiogenesis. *Pharmacology and Therapeutics*, 171, 56–64. https://doi.org/10.1016/j.pharmthera.2016.10.001

- Cho, J., D'Antuono, M., Glicksman, M., Wang, J., & Jonklaas, J. (2018). A review of clinical trials: mesenchymal stem cell transplant therapy in type 1 and type 2 diabetes mellitus. *American Journal of Stem Cells*, 7(4), 82–93. http://www.ncbi.nlm.nih.gov/pubmed/30510843
- De Sousa, P. A., Downie, J. M., Tye, B. J., Bruce, K., Dand, P., Dhanjal, S., Serhal, P., Harper, J., Turner, M., & Bateman, M. (2016). Development and production of good manufacturing practice grade human embryonic stem cell lines as source material for clinical application. *Stem Cell Research*, 17(2), 379–390. https://doi.org/10.1016/j. scr.2016.08.011
- Everett, E., & Mathioudakis, N. (2018). Update on management of diabetic foot ulcers. *Annals of the New York Academy of Sciences*, 1411(1), 153. https://doi.org/10.1111/nyas.13569
- Ezzatvar, Y., & García-Hermoso, A. (2022). Global estimates of diabetesrelated amputations incidence in 2010–2020: A systematic review and meta-analysis. *Diabetes Research and Clinical Practice*, *195*, Article 110194. https://doi.org/10.1016/j.diabres.2022.110194
- Freimark, D., Pino-Grace, P. P., Pohl, S., Weber, C., Wallrapp, C., Geigle, P., Pörtner, R., & Czermak, P. (2010). Use of encapsulated stem cells to overcome the bottleneck of cell availability for cell therapy approaches. *Transfusion Medicine and Hemotherapy*, 37(2), 66–73. https://doi. org/10.1159/000285777
- Fui, L. W., Lok, M. P. W., Govindasamy, V., Yong, T. K., Lek, T. K., & Das, A. K. (2019). Understanding the multifaceted mechanisms of diabetic wound healing and therapeutic application of stem cells conditioned medium in the healing process. *Journal of Tissue Engineering and Regenerative Medicine*, 13(12), 2218–2233. https://doi.org/10.1002/ term.2966
- Gardner, O. F. W., Fahy, N., Alini, M., & Stoddart, M. J. (2016). Differences in human mesenchymal stem cell secretomes during chondrogenic induction. *European Cells and Materials*, *31*, 221–235. https://doi. org/10.22203/eCM.v031a15
- Gnecchi, M., Danieli, P., Malpasso, G., & Ciuffreda, M. C. (2016). Paracrine mechanisms of mesenchymal stem cells in tissue repair. In M. Gnecchi (Ed.), *Methods in Molecular Biology volume 1416* (123–146). Humana Press. https://doi.org/10.1007/978-1-4939-3584-0_7

- González-González, A., García-Sánchez, D., Dotta, M., Rodríguez-Rey, J. C., & Pérez-Campo, F. M. (2020). Mesenchymal stem cells secretome: The cornerstone of cell-free regenerative medicine. *World Journal of Stem Cells*, 12(12), 1439–1690. https://doi.org/10.4252/wjsc.v12.i12.1529
- Hamzawy, M., Gouda, S. A. A., Rashid, L., Morcos, M. A., Shoukry, H., & Sharawy, N. (2017). The cellular selection between apoptosis and autophagy: Roles of vitamin D, glucose and immune response in diabetic nephropathy. *Endocrine*, 58, 66–80. https://doi.org/10.1007/ s12020-017-1402-6
- Hendrawan, S., Kusnadi, Y., Lagonda, C. A., Fauza, D., Lheman, J., Budi, E., Manurung, B. S., Baer, H. U., & Tan, S. T. (2021). Wound healing potential of human umbilical cord mesenchymal stem cell conditioned medium: An in vitro and in vivo study in diabetes-induced rats. *Veterinary World*, 14(8), 2109–2117. https://doi.org/10.14202/ vetworld.2021.2109-2117
- Hendrawan, S., Tan, S. T., & Budi, E. (2021). *Efek conditioned-medium human umbilical cord mesenchymal stem cell terhadap diabetes melitus pada tikus Sprague Dawley* [Unpublished data]. Fakultas Kedokteran, Universitas Tarumanagara.
- Hsiao, S. T. F., Asgari, A., Lokmic, Z., Sinclair, R., Dusting, G. J., Lim, S. Y., & Dilley, R. J. (2011). Comparative analysis of paracrine factor expression in human adult mesenchymal stem cells derived from bone marrow, adipose, and dermal tissue. *Stem Cells and Development*, 21(12), 2189–2203. https://doi.org/10.1089/scd.2011.0674
- Hu, J., Yu, X., Wang, Z., Wang, F., Wang, L., Gao, H., Chen, Y., Zhao, W., Jia, Z., Yan, S., & Wang, Y. (2013). Long term effects of implantation of WJ-MSCs for newly-onset type 1 diabetes mellitus. *Endocrine Journal*, 60(3), 347–357. https://doi.org/10.1507/endocrj.EJ12-0343
- Huang, P. C., Wang, G. J., Fan, M. J., Shibu, M. A., Liu, Y. T., Viswanadha, V. P., Lin, Y. L., Lai, C. H., Chen, Y. F., Liao, H. E., & Huang, C. Y. (2017). Cellular apoptosis and cardiac dysfunction in STZ-induced diabetic rats attenuated by anthocyanins via activation of IGFI-R/ PI3K/Akt survival signaling. *Environmental Toxicology*, 32(12), 2471– 2480. https://doi.org/10.1002/tox.22460
- Kim, H. O. H. S., Choi, S. M., & Kim, H. O. H. S. (2013). Mesenchymal stem cell-derived secretome and microvesicles as a cell-free therapeutics for neurodegenerative disorders. *Tissue Engineering and Regenerative Medicine*, 10(3), 93–101. https://doi.org/10.1007/s13770-013-0010-7

- Kim, S. J., Moon, G. J., Cho, Y. H., Kang, H. Y., Hyung, N. K., Kim, D., Lee, J. H., Nam, J. Y., & Bang, O. Y. (2012). Circulating mesenchymal stem cells microparticles in patients with cerebrovascular disease. *PLoS ONE*, 7(5), Article e37036. https://doi.org/10.1371/journal. pone.0037036
- Lai, R. C., Arslan, F., Tan, S. S., Tan, B., Choo, A., Lee, M. M., Chen, T. S., Teh, B. J., Eng, J. K. L., Sidik, H., Tanavde, V., Hwang, W. S., Lee, C. N., Oakley, R. M. El, Pasterkamp, G., de Kleijn, D. P. V., Tan, K. H., & Lim, S. K. (2010). Derivation and characterization of human fetal MSCs: An alternative cell source for large-scale production of cardioprotective microparticles. *Journal of Molecular and Cellular Cardiology*, 48(6), 1215–1224. https://doi.org/10.1016/j.yjmcc.2009.12.021
- Lai, R. C., Yeo, R. W. Y., & Lim, S. K. (2015). Mesenchymal stem cell exosomes. Seminars in Cell & Developmental Biology, 40, 82-88. https://doi.org/10.1016/J.SEMCDB.2015.03.001
- Li, L., Chen, X., Wang, W. E., & Zeng, C. (2016). How to improve the survival of transplanted mesenchymal stem cell in ischemic heart? *Stem Cells International.* https://doi.org/10.1155/2016/9682757
- Liang, X., Ding, Y., Zhang, Y., Tse, H. F., & Lian, Q. (2014). Paracrine mechanisms of mesenchymal stem cell-based therapy: Current status and perspectives. *Cell Transplantation*, 23(9), 1045–1059. https://doi. org/10.3727/096368913X667709
- Liu, X., Zheng, P., Wang, X., Dai, G., Cheng, H., Zhang, Z., Hua, R., Niu, X., Shi, J., & An, Y. (2014). A preliminary evaluation of efficacy and safety of Wharton's jelly mesenchymal stem cell transplantation in patients with type 2 diabetes mellitus. *Stem Cell Research & Therapy*, 5, Article 57. https://doi.org/10.1186/scrt446
- Lukomska, B., Stanaszek, L., Zuba-Surma, E., Legosz, P., Sarzynska, S., & Drela, K. (2019). Challenges and controversies in human mesenchymal stem cell therapy. *Stem Cells International*. https://doi. org/10.1155/2019/9628536
- Madrigal, M., Rao, K. S., & Riordan, N. H. (2014). A review of therapeutic effects of mesenchymal stem cell secretions and induction of secretory modification by different culture methods. *Journal of Translational Medicine*, 12, Article 260. https://doi.org/10.1186/s12967-014-0260-8
- Mauricio, D., Alonso, N., & Gratacòs, M. (2020). Chronic diabetes complications: the need to move beyond classical concepts. *Trends*

in Endocrinology and Metabolism, 31(4), 287–295. https://doi. org/10.1016/j.tem.2020.01.007

- Mitrousis, N., Fokina, A., & Shoichet, M. S. (2018). Biomaterials for cell transplantation. *Nature Reviews Materials*, *3*, 441–456. https://doi. org/10.1038/s41578-018-0057-0
- Mori, H. M., Kawanami, H., Kawahata, H., & Aoki, M. (2016). Wound healing potential of lavender oil by acceleration of granulation and wound contraction through induction of TGF- β in a rat model. *BMC Complementary and Alternative Medicine*, *16*, Article 144. https://doi. org/10.1186/s12906-016-1128-7
- Ohiagu, F. O., Chikezie, P. C., & Chikezie, C. M. (2021). Pathophysiology of diabetes mellitus complications: Metabolic events and control. *Biomedical Research and Therapy*, 8(3), 4243–4257. https://doi. org/10.15419/bmrat.v8i3.663
- Okonkwo, U. A., Chen, L., Ma, D., Haywood, V. A., Barakat, M., Urao, N., & DiPietro, L. A. (2020). Compromised angiogenesis and vascular Integrity in impaired diabetic wound healing. *PLoS ONE*, *15*(4), Article e0231962. https://doi.org/10.1371/journal.pone.0231962
- Okonkwo, U. A., & Dipietro, L. A. (2017). Diabetes and wound angiogenesis. *International Journal of Molecular Sciences*, *18*(7), Article 1419. https://doi.org/10.3390/ijms18071419
- Park, S. R., Kim, J.-W. W., Jun, H.-S. S., Roh, J. Y., Lee, H.-Y. Y., & Hong, I.-S. S. (2018). Stem cell secretome and its effect on cellular mechanisms relevant to wound healing. *Molecular Therapy*, 26(2), 606–617. https:// doi.org/10.1016/j.ymthe.2017.09.023
- Patel, S., Srivastava, S., Singh, M. R., & Singh, D. (2019). Mechanistic insight into diabetic wounds: Pathogenesis, molecular targets and treatment strategies to pace wound healing. *Biomedicine and Pharmacotherapy*, *112*, Article 108615. https://doi.org/10.1016/j.biopha.2019.108615
- Pawitan, J. A. (2014). Prospect of stem cell conditioned medium in regenerative medicine. *BioMed Research International*. https://doi. org/10.1155/2014/965849
- Peraturan Menteri Kesehatan Republik Indonesia (Permenkes) Nomor 32 Tahun 2018 tentang Penyelenggaraan Pelayanan Sel Punca dan/atau Sel. (2018). https://peraturan.bpk.go.id/Details/111942/permenkes-no-32-tahun-2018
- Pinho, A. G., Cibrão, J. R., Silva, N. A., Monteiro, S., & Salgado, A. J. (2020). Cell secretome: Basic insights and therapeutic opportunities

for CNS disorders. *Pharmaceuticals*, 13(2), Article 31. https://doi. org/10.3390/ph13020031

- Saheli, M., Bayat, M., Ganji, R., Hendudari, F., Kheirjou, R., Pakzad, M., Najar, B., & Piryaei, A. (2020). Human mesenchymal stem cellsconditioned medium improves diabetic wound healing mainly through modulating fibroblast behaviors. *Archives of Dermatological Research*, 312(5), 325–336. https://doi.org/10.1007/s00403-019-02016-6
- Shi, G. J., Li, Z. M., Zheng, J., Chen, J., Han, X. X., Wu, J., Li, G. Y., Chang, Q., Li, Y. X., & Yu, J. Q. (2017). Diabetes associated with male reproductive system damages: Onset of presentation, pathophysiological mechanisms and drug intervention. *Biomedicine and Pharmacotherapy*, 90, 562–574. https://doi.org/10.1016/j.biopha.2017.03.074
- Shi, G. J., Shi, G. R., Zhou, J., Zhang, W., Gao, C., Jiang, Y., Zi, Z. G., Zhao, H., Yang, Y., & Yu, J. Q. (2018). Involvement of growth factors in diabetes mellitus and its complications: A general review. *Biomedicine* and Pharmacotherapy, 101, 510–527. https://doi.org/10.1016/j. biopha.2018.02.105
- Shin, S. H., Koh, Y. G., Lee, W. G., Seok, J., & Park, K. Y. (2022). The use of epidermal growth factor in dermatological practice. *International Wound Journal*, 20(6), 2414–2423. https://doi.org/10.1111/iwj.14075
- Shin, S., Lee, J., Kwon, Y., Park, K. S., Jeong, J. H., Choi, S. J., Bang, S. I., Chang, J. W., & Lee, C. (2021). Comparative proteomic analysis of the mesenchymal stem cells secretome from adipose, bone marrow, placenta and Wharton's jelly. *International Journal of Molecular Sciences*, 22(2): Article 845. https://doi.org/10.3390/ijms22020845
- Sortwell, C. E., Pitzer, M. R., & Collier, T. J. (2000). Time course of apoptotic cell death within mesencephalic cell suspension grafts: Implications for improving grafted dopamine neuron survival. *Experimental Neurology*, 165(2), 268–277. https://doi.org/10.1006/exnr.2000.7476
- Tan, S. T., Aisyah, P. B., Firmansyah, Y., Nathasia, N., Budi, E., & Hendrawan, S. (2023). Effectiveness of secretome from human umbilical cord mesenchymal stem cells in gel (10 % SM-hUCMSC Gel) for chronic wounds (diabetic and trophic ulcer) – phase 2 clinical trial. *Journal of Multidisciplinary Healthcare*, 16, 1763–1777. https://doi.org/10.2147/ JMDH.S408162
- Tan, S. T., Hendrawan, S., Dewi, A. K., Nuraeni. (2021). Efek anti-inflamasi dan antioksidan pemberian conditioned media human umbilical cord-

mesenchymal stem cells (CM hUC-MSC) intravena pada tikus Sprague Dawley normal [Unpublished data]. Fakultas Kedokteran, Universitas Tarumanagara.

- Teixeira, F., & Salgado, A. (2020). Mesenchymal stem cells secretome: Current trends and future challenges. *Neural Regeneration Research*, 15(1), 75–77. https://doi.org/10.4103/1673-5374.264455
- Velnar, T., & Gradisnik, L. (2018). Tissue augmentation in wound healing: The role of endothelial and epithelial cells. *Medical Archives (Sarajevo, Bosnia and Herzegovina)*, 72(6), 444–448. https://doi.org/10.5455/ medarh.2018.72.444-448
- Vizoso, F., Eiro, N., Cid, S., Schneider, J., & Perez-Fernandez, R. (2017). Mesenchymal stem cell secretome: Toward cell-free therapeutic strategies in regenerative medicine. *International Journal of Molecular Sciences*, 18(9), Article 1852. https://doi.org/10.3390/ijms18091852
- Wang, B., Pang, M., Song, Y., Wang, H., Qi, P., Bai, S., Lei, X., Wei, S., Zong, Z., Lin, S., Zhang, X., Cen, X., Wang, X., Yang, Y., Li, Y., Wang, Y., Xu, H., Huang, L., Tortorella, M., ... Li, G. (2023). Human fetal mesenchymal stem cells secretome promotes scarless diabetic wound healing through heat-shock protein family. *Bioengineering and Translational Medicine*, 8(1), Article e10354. https://doi.org/10.1002/ btm2.10354
- Wang, M., Yuan, Q., & Xie, L. (2018). Mesenchymal stem cell-based immunomodulation: Properties and clinical application. *Stem Cells International*. https://doi.org/10.1155/2018/3057624
- Wang, Y., Yi, H., & Song, Y. (2021). The safety of MSC therapy over the past 15 years: A meta-analysis. *Stem Cell Research and Therapy*, *12*, Article 545. https://doi.org/10.1186/s13287-021-02609-x
- Xia, J., Minamino, S., Kuwabara, K., & Arai, S. (2019). Stem cell secretome as a new booster for regenerative medicine. *BioScience Trends*, 13(4), 299–307. https://doi.org/10.5582/bst.2019.01226
- Xiang, E., Han, B., Zhang, Q., Rao, W., Wang, Z., Chang, C., Zhang, Y., Tu, C., Li, C., & Wu, D. (2020). Human umbilical cord-derived mesenchymal stem cells prevent the progression of early diabetic nephropathy through inhibiting inflammation and fibrosis. *Stem Cell Research and Therapy*, 11, Article 336. https://doi.org/10.1186/s13287-020-01852-y

- Zaghary, W. A., Elansary, M. M., Shouman, D. N., Abdelrahim, A. A., Abu-Zied, K. M., & Sakr, T. M. (2021). Can nanotechnology overcome challenges facing stem cell therapy? A review. *Journal of Drug Delivery Science and Technology*, 66, Article 102883. https://doi.org/10.1016/j. jddst.2021.102883
- Zhao, L., Hu, C., Zhang, P., Jiang, H., & Chen, J. (2019). Preconditioning strategies for improving the survival rate and paracrine ability of mesenchymal stem cells in acute kidney injury. *Journal of Cellular* and Molecular Medicine, 23(2), 720–730. https://doi.org/10.1111/ jcmm.14035
- Zhao, Q., Ren, H., & Han, Z. (2016). Mesenchymal stem cells: Immunomodulatory capability and clinical potential in immune diseases. *Journal of Cellular Immunotherapy*, 2(1), 3–20. https://doi. org/10.1016/j.jocit.2014.12.001



Saba Majeed

Saba Majeed is an Assistant Professor of Pharmacology, Faculty of Pharmacy, Ziauddin University. She graduated from the field of Pharmacology and Neuroscience from the International Center for Chemical and Biological Sciences, Faculty of

Pharmacy, University of Karachi. Her primary interest is neural stem cell research, which aims to modulate neurogenesis to treat neurodegenerative disorders. She has a great passion for exploring novel technologies and therapeutic treatment options to control neurological deficit-related orders; she has worked on neurogenesis and is involved in establishing novel models of co-culturing stem cells and neurons. Her open and contextual evaluation model is based on co-culturing techniques and promotes neurogenesis by exploring new signaling pathways for improving healthcare. She has patented her Ph.D research in neurogenesis for US Patent Isoxylitones mediated neurogenesis. Currently, she works as an assistant professor at Ziauddin University's Department of Pharmacology and is involved in research area of neuroinflammation targeting various signaling mechanisms modulating neurodegenerative diseases like alzhiemer's, stroke, parkinson's, etc. She has an excellent command on all molecular and imaging techniques. Email: saba.majeed@zu.edu.pk



Siufui Hendrawan

Siufui Hendrawan is an Associate Professor at the Faculty of Medicine, Tarumanagara University, Jakarta, Indonesia. She is also the head of Tarumanagara Human Cell Technology (THCT) Laboratory, a research laboratory founded in 2011 in collaboration with Baermed, Switzerland. She completed her PhD at Hassanuddin University, Makassar, Indonesia, in 2017 and has published more than 15 papers in national and international journals. She specializes in tissue engineering, cell biology, and biomaterials science. She has conducted numerous preclinical studies and some clinical trials. Her research works specialize in tissue engineering, cell biology, and biomaterials science. She pioneers research focused on the development of mini-organs and cell-matrix implants, as well as the therapeutic application of secretome from mesenchymal stem cells. Email: siufui@fk.untar.ac.id



Somia Gul

Somia Gul is a professor at Department of Pharmaceutical Chemistry Faculty of Pharmacy, Jinnah University for Women. She has completed her specialization in Pharmaceutical Chemistry and earned her Ph.D as an HEC Indigenous Ph.D fellowship Scholar from the University of Karachi in 2011. Somia has over 17 years of teaching, research, and industrial experience as a

supervisor of various M. Phil. and PhD. research scholars, research projects, and training sections. She has more than 103 research publications in reputed journals with good impact factors and one book published.

Her specialization is in medicinal chemistry, and her expertise and research interests mainly focus on drug-drug interactions of different classes of drugs, organic derivative synthesis and metal complexes, natural product chemistry, formulation development, validation and biological evaluation of herbal origin, methods development, and validation for analysis of drugs through HPLC and UV/Vis. Spectroscopy.

Somia's current research focuses on in-silico drug design, synthesis, biological evaluation, and molecular docking studies of novel analogues of biological interest, especially antimicrobials "Discovering the Miracles of Stem Cells" offers an exhaustive exploration of stem cell research, underscoring the profound transformative potential of these cells across various medical and scientific domains. This scholarly work meticulously delineates the fundamental characteristics of stem cells, including their types, functions, and pivotal roles in tissue regeneration and therapeutic interventions. The text is thoughtfully structured into multiple chapters, each focusing on distinct applications and areas of research.

The opening chapter provides a detailed exposition of the culture techniques for mesenchymal stem cells (MSCs), crucial for advances in regenerative medicine. Subsequent chapters delve into the application of stem cells in orthopedics, showcasing their effectiveness in treating bone and cartilage disorders. Further discussions cover the use of MSCs in healing diabetic wounds, the advantages of MSC-conditioned mediums in combating skin aging, and a critical evaluation of the safety and efficacy of stem cells in managing acute myocardial infarction. A particular emphasis is placed on the potential of CD34+ hematopoietic stem cells in skin rejuvenation following UV exposure, supported by robust empirical research on animal models. Later sections explore the therapeutic implications of stem cells in neurological disorders, highlighting their utility in managing neurodegeneration and other cerebral conditions. Additionally, the book examines the ethical dimensions of stem cell research, advocating for stringent ethical standards to navigate the complex moral landscapes encountered in the use of stem cells for therapeutic purposes. It also addresses the unique opportunities and challenges associated with stem cell research and application in Indonesia, reflecting on the country-specific context that shapes these endeavors.

Overall, this text serves as a comprehensive resource that elucidates both the scientific and therapeutic dimensions of stem cells while addressing the ethical, regulatory, and practical challenges in the field. Designed to equip readers with a thorough understanding of how stem cells can be utilized to enhance health outcomes and treat a wide array of diseases, this book positions stem cell research as a beacon of hope within the medical and biotechnological domains, yet acknowledges the intricate ethical, technical, and regulatory challenges that must be navigated with precision and prudence.



Published by: BRIN Publishing, member of Ikapi Gedung B.J. Habibie Lt. 8, Jin. M.H. Thamrin No. 8, Jakarta Pusat 10340 E-mail: penerbit@brin.go.id Website: penerbit.brin.go.id



